A somatic mutation in the thyrotropin receptor gene in a patient with an autonomous nodule within a multinodular goiter


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ABSTRACT

Thyrotropin (TSH) is the prime regulator of thyroid cell growth and function and acts through the thyrotropin receptor (TSHR) located on the surface membrane of thyrocytes. Somatic heterozygous mutations that cause TSHR activation in the absence of TSH have been found in toxic adenomas and in hot nodules of multinodular goiters. Clinically and histologically heterogeneous nodules can share common gain-of-function mutations. Mutation prevalence varies greatly and is inversely related to iodine intake of the population. We report a Greek patient presenting with subclinical hyperthyroidism due to a fast-growing autonomous hyperplastic nodule in a long-standing multinodular goiter. Direct DNA sequencing showed that the hot nodule harbored a somatic heterozygous activating TSHR mutation: substitution of glutamine for leucine in the third transmembrane helix. This mutation (L512Q) was recently described in two solitary toxic adenomas. This report expands the spectrum of mutations shared by dissimilar hot nodules, supporting a common mechanism for nonautoimmune thyroid autonomy. The identification of the L512Q substitution demonstrates that gain-of-function TSHR mutations are encountered in Greece, although iodine deficiency has been significantly corrected over the last three decades.

Key words: Thyrotropin Receptor gene, autonomous thyroid nodules, TRHR gene mutations

INTRODUCTION

Under physiological conditions, differentiation and proliferation of thyroid cells are regulated primarily by thyrotropin (TSH). TSH acts by binding to the TSH receptor (TSHR) on thyrocyte membranes. TSHR belongs to the family of G protein-coupled receptors, with the typical motif of seven transmembrane segments connected by three extra- and three intracellular loops (Figure 1). Thyrotropin binding to TSHR activates, through Gsα protein, the cAMP cascade mainly responsible for mediating TSH effects. At higher hormone concentrations,
the less important inositol phosphate pathway is also trig-
gerated via Gsα.

It was predicted that somatic gain-of-function mutations affecting proteins of the TSH signalling pathway would lead to the clonal expansion of the mutated cell and the generation of a clone of autonomously growing and functioning thyrocytes. This would correspond to the clinical entity of solitary toxic adenoma. Activating mutations were indeed identified in toxic adenomas, initially in the Gsα protein and later in the TSHR itself. Mutations are somatic, heterozygous and confined to the autonomous tissue; all are located in exons 9 and 10 of the receptor gene, which encode for the intracellular, transmembrane and proximal extracellular domains. No activating mutations were found within TSHR exons 1-8, which encode for most of the extracellular domain. Mutant receptors cause constitutive (TSH-independent) activation of the cAMP and/or inositol phosphate pathways. The prevalence of gain-of-function TSHR mutations is highly variable among different series. Methodological discrepancies are partly incriminated, but different iodine intake of populations also seems responsible.

Constitutively activating TSHR mutations were subsequently found in solitary or multiple hot nodules of toxic or autonomous multinodular goiters. Toxic multinodular goiter represents the final phase in the evolution of goiter over time; it develops slowly within a gland whose nodules gain autonomy. In this clinical setting, autonomous nodules are histologically classified as either typical adenomas or hyperplastic nodules and gain-of-function mutations exist in both types. Although not present in all hot nodules, these mutations emerge as a possible common mechanism underlying the pathogenesis of non-autoimmune thyroid autonomy.

We report here the first activating mutation identified in a hyperplastic hot nodule of a Greek patient with autonomous multinodular goiter. By direct genomic sequencing of the TSHR exon 10 we found a single aminoacid substitution, L512Q, in the third transmembrane segment of the receptor (Figure 1). This mutation was recently identified by different methodology in two German patients with solitary toxic adenomas.

**PATIENT REPORT**

A 50-year old female patient with a 10-year history of euthyroid multinodular goiter was seen in the outpatient endocrine clinic. She was on no medication. Her main complaint was a recent swelling of the right lobe of the thyroid gland. She was clinically euthyroid. Laboratory investigation showed a T3 value of 2.4 nmol/L (160ng/dL) (normal range: 1.2-3.0 nmol/L, 80-200 ng/dL), a T4 value of 92 nmol/L (7.2 µg/dL) (normal range: 58-160 nmol/L, 4.5-12.5 µg/dL) and a TSH value of 0.35 mU/L (0.35 μU/mL) (normal range: 0.4-4.0), indicating subclinical hyperthyroidism. The antiperoxidase-(TPO) and antithyroglobulin antibodies were negative. Thyroid ultrasound showed multiple nodules bilaterally within an enlarged thyroid gland (right lobe: 5.8x1.6x2.2 cm, left lobe: 4.5x1.5x1.3 cm). The dominant nodule was in the inferior pole of the right lobe, cystic in consistency and 3.5 cm in diameter. A 99m-Tc scintiscan revealed increased uptake by the dominant nodule and suppression of the remaining parenchyma. The patient was subjected to subtotal thyroidectomy. Histologically, all thyroid nodules showed a hyperplastic pattern.

Immediately after thyroidectomy, tissue samples were taken from the autonomous nodule and surrounding normal tissue. Samples were carefully excised matching the scintiscan and ultrasound patterns with the whole gland laid in its proper anatomic orientation. Tissue specimens were shock-frozen in liquid nitrogen. Peripheral blood sample was collected in EDTA and stored at -20°C.

The study complied with the stipulations of the dec-
laration of Helsinki. Informed consent was obtained from
the patient before surgery.

Materials and Methods

Genomic DNA was extracted from surgical samples
following the standard phenol/chlorophorm procedure
and from peripheral blood leukocytes using Nucleospin
Blood QuickPure kit (Macherey-Nagel, Düren, Ger-
many). The complete exon 10 of the thyrotropin recep-
tor gene, which encodes for the transmembrane and in-
tracellular domains of TSHR, was amplified by polymer-
ase chain reaction (PCR) as previously described15. Two
overlapping sequences were amplified with two sets of
oligonucleotide primers (MWG-Biotech AG, Ebersberg,
Germany): An 875bp fragment with forward primer 5/-
ACTGTCTTTGCAAGCGAGTT- 3/and reverse prim-
er 5/-GTGTCATGGGATTTGGAATGC-3/ and an 868bp
fragment with forward primer 5/-ACTGTCTTT-
GCAAGCGAGTT -3/ and reverse primer 5/ -GTGT-
CATGGGATTTGGAATGC- 3/. 100 ng of genomic DNA
was used as template in a PTC-200 thermocycler (MJ
Research Inc., Waltham, MA, USA). PCR was per-
formed in a total volume of 50 µl, using 10 pmol of each
appropriate primer, 200 µM of each dNTP, 1.5 mM MgCl,
2.5 U Gibco Brl Taq DNA polymerase (ANTI-SEL,
Thessaloniki, Greece) and 10x reaction buffer contain-
ing 200mM Tris-HCl (pH=8.4) and 500 mM KCl. After
an initial denaturing at 95°C for 5 min, reactions were
subjected to 30 cycles of 30 sec denaturing at 95°C, an-
nealing at 56°C and extension at 72°C, followed by a
final 6 min extension at 72°C. PCR products were puri-
ified with Nucleospin Extract kit (Macherey-Nagel,
Düren, Germany). At least two different purified PCR
amplicons were sequenced on both sense and anti-sense
strands. Sequencing reactions were performed using ABI
PRISM BigDye Terminator Cycle Sequencing Ready
Reaction kit (Applied Biosystems, Weiterstadt, Ger-
rnany) with the PCR primers as sequencing primers and run
on an ABI 373 DNA Sequencer (Applied Biosystems,
Foster City, CA, USA).

Results

Direct automated sequencing of thyrotropin receptor
(TSHR) exon 10 showed a T to A transversion in the
heterozygous state (Figure 2) in the hyperfunctioning
hyperplastic nodule. This result indicated the substitu-
tion of glutamine (CAG) for leucine (CTG) at codon 512
in the third transmembrane helix (Figure 1). The height of
the mutant and wild-type base signal peaks was almost
identical (Figure 2). This indicates that the sample
consisted purely of tissue heterozygous for the L512Q
mutation. The substitution was absent from surrounding
non-autonomous tissue and peripheral blood leukocytes
(Figure 2), proving that the mutation was somatic. This
is a gain-of-function mutation recently described in two
German patients harboring solitary toxic adenomas14.
When transfected in COS7 cells14, the mutant receptor
reached 87% of the wild-type TSHR cell surface expres-
sion level. It promoted a basal cAMP accumulation five
times higher than wild-type receptor and a TSH-stimu-
lated response slightly less than wild-type.

Discussion

Somatic heterozygous gain-of-function mutations of
the thyrotropin receptor (TSHR) and Gsα protein were
Initially described in solitary toxic adenomas of the thyroid gland\(^4,5\). These mutations cause TSH-independent (constitutive) activation of the cAMP and/or inositol phosphate signalling pathways\(^6\), which control thyrocyte growth and function\(^7\). Activating mutations were subsequently identified in hot nodules of toxic or autonomous multinodular goiters\(^9,10\). The term "toxic multinodular goiter" (TMG) encompasses a wide spectrum of different clinical entities, ranging from a single hyperfunctioning nodule within an enlarged thyroid gland having additional nonfunctioning nodules, to multiple hyperfunctioning areas scattered throughout the gland, barely distinguishable from nonfunctioning nodules and extranodular parenchyma\(^11\). Moreover, hyperfunctioning nodules may present histologically as typical adenomas (encapsulated homogeneous follicular structures) or as hyperplastic nodules (micro- macrofollicular aggregates devoid of capsule)\(^11\). Constitutively activating TSHR and Gs\(\alpha\) mutations have been found in autonomous tissues of all these clinical and pathologic variants\(^12,13\). The L512Q mutation was recently reported in two German patients with toxic uninodular goiter, whose hot nodules showed the typical histological presentation of adenoma\(^14\). We have identified the same mutation in a hyperplastic hot nodule of a patient with autonomous multinodular goiter. This expands the spectrum of mutations shared by autonomous nodules of different clinical and histological backgrounds. Thus, it reinforces the concept of a common mechanism underlying the pathogenesis of non-autoimmune autonomy\(^6\).

Functional studies of the L512Q TSHR revealed a constitutive activation of the cAMP cascade five-fold over the wild-type receptor\(^14\). Although the signalling through the PIP2 phospholipase-C cascade was not studied, in comparison to other activating TSHR mutations\(^17\), this represents a relatively strong constitutive activity. Our patient presented with subclinical hyperthyroidism due to a recently growing nodule in a long-standing multinodular goiter. This is in accordance with L512Q driven stimulation of thyrocyte growth and function. However, the clinical characteristics of patients with autonomous nodules harboring activating mutations are not necessarily associated with receptor constitutive activity\(^6\). Identical somatic or germline mutations can present with highly variable clinical pictures\(^6\). This implies that other factors are involved in the clinical expression of an activating event in the TSHR signalling pathway\(^2\), including alterations of the G-protein signalling or overexpression of the TSHR itself due to defects in receptor desensitization and internalization\(^18\).

Interestingly, in the two adenomas previously reported to harbour the L512Q substitution\(^14\), initial direct sequencing of the nodules’ genomic DNA failed to identify the mutation. The mutation was subsequently found by direct sequencing of heteroduplex bands formed in a denaturing gradient gel electrophoresis (DGGE). This demonstrated the ability of DGGE to recognize the presence of heterozygous mutations against a higher background of wild type sequence\(^19\). This is important when heterozygous somatic mutations are sought, because tissue samples may contain fibroblasts, blood leukocytes, or surrounding normal tissue. Such contamination decreases the percentage of the mutant allele in the sample below the ideal 50%, and possibly below the detection limit of direct automated sequencing. This has been advocated as one of the reasons for the highly variable prevalence of TSHR and Gs\(\alpha\) mutations in different series\(^7\). The identical height of the mutant and normal base peaks (Figure 2), indicates the absence of contamination in our sample. This emphasizes the importance of meticulous sample selection in conjunction with a sensitive detection method.

Codon 512 in the third transmembrane helix (TM3) of TSHR emerges as a receptor site frequently mutated in autonomous nodules. L512Q has been found in three cases, including this report\(^14\). Another activating substitution at the same codon, L512R, has been reported twice\(^14,20\). TM3 encompasses the TSHR highly conserved aminoacids 497-517. Naturally occurring activating mutations in TM3 also affect aminoacids 505 and 509. In vitro mutagenesis of residues 505-513 demonstrated that mutations at positions other than 505, 509 and 512 do not induce constitutive receptor activation\(^29\). These three aminoacids are presumably located on the surface of a simple \(\alpha\)-helix, facing in the same direction. They may be involved in formation of non-covalent bonds with another helix that participate in maintaining the receptor in a constrained inactive conformation\(^29\). In contrast to initial reports identifying mutations within or near the third intracellular loop\(^5\), it is now evident that activating mutations are scattered throughout the carboxyl half of the receptor\(^6\). This emphasizes the need for screening completely the corresponding exon 10 of the TSHR gene. The identification of frequently affected residues provides insight into the TSHR structure-function relationship and the mechanisms of receptor activation.

Gain-of-function mutations have been reported with varying frequencies in different series of solitary toxic adenomas, ranging from 0% to 82%\(^7\). Methodological reasons may partly account for such extreme discrepancies. Interestingly, the prevalence of activating mutations...
is inversely related to iodine intake of the population, as is the prevalence of TMG. Here we report the first TSHR mutation found in a Greek patient with sub-toxic multinodular goiter. Iodine deficiency was well documented in post-war Greece, but has been corrected over the last 30 years due to increased use of iodized salt and improved socioeconomic conditions. Although iodine deficiency still persists in small mountainous regions, particularly in areas of Southwestern Greece, the prevalence of TMG is inversely related to iodine intake of the population.

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